

Sodium alginate for wound healing applications: A review

Muhammad Tahir^{1,A–D}, Simone Pettineo^{2,A,B}, Silvia Vicini^{2,A,E,F}, Maila Castellano^{2,E,F}, Marina Alloisio^{2,E,F}, Ketul Popat^{3,E,F}, Harishkumar Madhyastha^{4,E,F}, Kaoru Ohe^{5,E,F}, Kentaro Sakai^{6,E,F}, Alina Sionkowska^{1,A,E,F}

¹ Laboratory for Biomaterials and Cosmetics, Faculty of Chemistry, Nicolaus Copernicus University in Torun, Poland

² Department of Chemistry and Industrial Chemistry, University of Genova, Italy

³ Department of Bioengineering, George Mason University, Fairfax, USA

⁴ Department of Cardiovascular Physiology, Faculty of Medicine, University of Miyazaki, Japan

⁵ Center for Science and Engineering Education, Faculty of Engineering, University of Miyazaki, Japan

⁶ Division for Research Facility Support Organization for Promotion of Research and Industry – Academic Regional Collaboration, University of Miyazaki, Japan

A – research concept and design; B – collection and/or assembly of data; C – data analysis and interpretation;

D – writing the article; E – critical revision of the article; F – final approval of the article

Polymers in Medicine, ISSN 0370-0747, eISSN 2451-2699

Polim Med. 2026;56(1):7–17

Address for correspondence

Muhammad Tahir

E-mail: 503438@o365.doktorant.umk.pl

Funding sources

None declared

Conflict of interest

None declared

Acknowledgements

None declared

Received on August 1, 2025

Reviewed on September 27, 2025

Accepted on February 26, 2026

Published online on July 30, 2026

Abstract

Sodium alginate (SA) is a polysaccharide biopolymer widely used in wound healing applications due to its beneficial role in the hemostatic, inflammatory, and proliferative phases of tissue repair. Its hydrophilic nature supports the wound healing process by enabling efficient absorption of wound exudate and maintaining a moist microenvironment conducive to tissue regeneration. Moreover, SA can form highly porous structures that promote oxygen diffusion and provide a suitable scaffold for neovascularization and new tissue formation. This review summarizes recent advances in the application of SA in wound dressings. A bibliometric analysis of Scopus data using the keywords “sodium alginate” AND “wound healing” reveals a growing number of publications in recent years, highlighting the increasing scientific interest in this field. The expanding utilization of SA in wound healing may be attributed to its favorable properties, including biocompatibility, biodegradability, and low toxicity.

Key words: wound healing, sodium alginate, biopolymer, alginic acid, porous materials

Cite as

Tahir M, Pettineo S, Vicini S, et al. Sodium alginate for wound healing applications: A review. *Polim Med.* 2026;56(1):7–17. doi:10.17219/pim/218616

DOI

10.17219/pim/218616

Copyright

Copyright by Author(s)

This is an article distributed under the terms of the Creative Commons Attribution 3.0 Unported (CC BY 3.0) (<https://creativecommons.org/licenses/by/3.0/>)

Introduction

Biopolymers can be extracted from bio-based materials using various techniques. They can also be produced biotechnologically or synthesized directly from building blocks derived from living organisms. Thus, in general, nature is the primary source of biopolymers.¹ Jabeen and Atif defined biopolymers as macromolecules that are primarily composed of monomeric units of natural origin.² Biopolymers are advantageous due to their environmentally friendly nature.³ Nitta and Numata discussed the classification of biopolymers, which are mainly divided into 3 types: polysaccharides, nucleic acids, and proteins.⁴ In the huge family of polysaccharides, one can find starch, cellulose, chitin, chitosan, and many more, for example, alginate.⁵

The primary sources of alginate reported are brown seaweed and marine algae.^{6,7} Alginate has a unique non-toxic nature and biocompatibility, which make this biopolymer attractive for biomedical and cosmetic applications.^{8,9} It exhibits bioadhesivity, which is the primary reason for its widespread use in pharmaceutical applications.¹⁰ It may form sponge-like structures that are capable of absorbing water.¹¹ Alginate-based hydrogels are formed through ionic crosslinking.¹² Salarvand et al. reported the presence of hydroxyl and carboxyl groups in alginate, which gives the possibility of hydrogen bond formation.¹³ Moreover, alginate contains negative ions capable of further interactions.¹⁴ Wang et al. reported that sodium alginate (SA) and its oxidized derivatives show good cytocompatibility.¹⁵ Alginate-based nanofibers are widely utilized in biomedical applications, especially in tissue engineering and wound healing.^{16,17} It has recently been observed that SA applications in various areas are growing, particularly in wound healing.

Sodium alginate

The properties of SA are widely studied. Phùng et al. mentioned the water solubility of SA,¹⁸ whereas Yan et al. reported that SA is insoluble in ethanol.¹⁹ Sodium alginate is highly preferred in several applications due to its outstanding ability to form films.^{20,21} Yan et al. described SA as a low-cost polymer presenting high potential.¹⁹

The presence of functional groups, such as hydroxyl and carboxyl groups, makes it easy to modify.²² Belalia and Djelali reported the rheological behavior of SA. In their research, it was concluded that it presents shear-thinning non-Newtonian flow behavior in its aqueous form.²³ Viscosity was reduced with an increase in shear rate; this behavior has been reported as shear-thinning behavior in the literature.²⁴

In the structural formula of SA, the main constituents are α -L-guluronic acid and β -D-mannuronic acid.^{25,26} ChemDraw 25.0.2 software (Revvity Signals Software) was used to draw the structural formulas of SA and its constituents, which are presented in Fig. 1. The “Structure” tab of the ChemDraw software was selected, and in this tab, “Convert Name to Structure” was chosen to draw the structural formulas of SA and its constituents. The names used are also presented in the figure alongside their corresponding structural formulas. Therefore, in short, SA is a combination of 2 uronic acids. Sodium alginate sourced from algae contains a significant portion of other cations: Ca (calcium), Mg (magnesium), and Sr (strontium), which are treated with hydrochloric acid (HCl) to obtain alginic acid through a liquid wash mechanism.²⁷

The addition of sodium carbonate (Na_2CO_3) leads to a pH range of 9–10. Na_2CO_3 can also be replaced with alternatives such as sodium hydroxide (NaOH), sodium bicarbonate (NaHCO_3), or sodium chloride (NaCl). This leads to the conversion of alginic acid (which is water-insoluble) to SA (a water-soluble form).²⁸ It can be followed by either the Ca-alginate or the alginic acid route. The route considering alginic acid involves the treatment of SA with HCl. The 2nd route is the Ca-alginate route, which involves the addition of calcium chloride to SA to form Ca-alginate.²⁹ Ca-alginate is further treated with HCl to obtain alginic acid. The alginic acid obtained from these 2 routes is further treated with Na_2CO_3 to produce SA in its pure form.³⁰

Porosity of sodium alginate materials

Sodium alginate materials can be fabricated into gels. Abka-Khajouei et al. reported that alginate gels presented pores, with the pore size specified in their research ranging

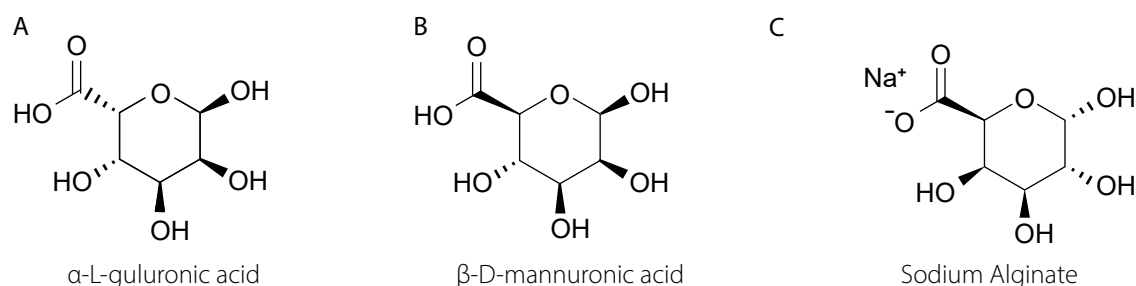


Fig. 1. Structural formulas of (A) α -L-guluronic acid, (B) β -D-mannuronic acid, and (C) sodium alginate

from 5 nm to 200 nm. They mentioned electron microscopy for pore size demonstration.³¹ Da Silva et al. reported that as the SA concentration increases, it causes a decrease in the material's porosity. Therefore, an inverse relationship between SA concentration and material porosity was presented. Porosity in wound dressings is preferred because it facilitates the process of oxygenation.³² Sinha et al. reported that wound dressings with the highest porosity had the greatest tendency to absorb wound exudate.³³ Liu et al.³⁴ also demonstrated that SA gel presented pores. They conducted field emission electron microscopy (FE-SEM) on SA gels with the addition of calcium ions (Ca^{2+}) to demonstrate the pores.

MATLAB and Python are referred to as programming languages.^{35,36} They are effectively utilized for the visualization and demonstration of pores. Jenkins et al. were the developers of PoreScript (a MATLAB algorithm), and PoreScript was used to determine pore size from scanning electron microscopy (SEM) images.³⁷ Polez et al. utilized SEM images to determine porosity using PoreSpy,³⁸ a Python package that characterizes porous materials from 3D images.³⁹ Negut and Bitu reported the use of a convolutional neural network (CNN) to identify pore size distribution in hydrogels.⁴⁰ Zhang et al. reported CNN as a deep learning model.⁴¹ El Naqa and Murphy described deep learning as a sub-classification of machine learning, and stated that deep learning has the capacity to be trained from raw data.⁴² Machine learning has been referred to as a field within artificial intelligence.⁴³ Three types of machine learning are reported in the literature: supervised learning, unsupervised learning, and reinforcement learning.^{44,45}

Supervised machine learning utilizes labeled data.⁴⁶ Unsupervised learning is based on unlabeled data.⁴⁷ Reinforcement learning allows an agent to take actions.⁴⁸ Correct actions in reinforcement learning lead to earning rewards, whereas wrong decisions result in failure.⁴⁹ Saberian et al. utilized ImageJ software (National Institutes of Health (NIH), Bethesda, USA) to measure pores based on SEM images. The hydrogel utilized in their research was based on alginate/chitosan and also included honey and aloe vera. The prepared hydrogel was intended for use as a wound dressing.⁵⁰ Olevsky et al. developed PoreVision using a Python script. PoreVision utilizes Python libraries, primarily OpenCV, NumPy, and Pandas. It was used to measure pore size from SEM images and can be utilized for gels and porous materials, as indicated in their research.⁵¹ Shkarin et al. developed quanfima, a Python-supported package that can be used to analyze the morphology of biomaterials, including porosity.⁵² MICPY has been highlighted for pore analysis and has been reported as a Python-supported package. It can analyze SEM images.^{53,54} Nair et al. utilized CTAnalyzer software for pore size determination, which was made possible by using a 3D object analyzer within the software.⁵⁵ Karaca and Aldemir Dikici utilized a deep learning model, which they named Pore D². In their study, YOLOv5 was used for

object detection, and SEM images were utilized as the input. EasyOCR has been used to extract text from images, and it is a Python library that employs a deep learning approach. The output layer provided accurate information on pore size.⁵⁶

Wettability of sodium alginate materials

The hydrophilicity of the material's surface may be a reason for better cell adhesion. Yuan and Lee presented the relationship between the water contact angle and wettability in the context of liquid–solid interaction. According to their research, contact angles below 90° exhibited the highest wettability. A contact angle greater than 90° was a clear indication of low wettability.⁵⁷ Kumar and Prabhu described reactive and non-reactive wetting. A reaction occurring between the spreading liquid and the substrate material impacts the mechanism of wetting, which is reported as reactive wetting. If no reaction occurs between the spreading liquid and the substrate material, it is reported as inert or non-reactive wetting.⁵⁸ Xie et al. conducted contact angle analysis for membranes made of SA, and SA 100 and SA 200 were named for the lower amounts of guluronic acid. Manugel DMB was named for a higher amount of guluronic acid. A Milli-Q water drop was utilized to obtain contact angle measurements, and the analysis was conducted on alginate–chitosan–alginate (ACA) membranes. The water contact angle for SA 100, as reported in their study, was $65.9 \pm 1.0^\circ$. SA 200 presented a water contact angle of $68.5 \pm 2.4^\circ$. The water contact angle of Manugel DMB recorded in their research was $77.7 \pm 3.3^\circ$. These values are reported as mean \pm standard error (SE). The number of analyses specified in their study was 3. In short, their results showed that an increase in guluronic acid content led to a reduction in hydrophilicity.⁵⁹ Karmakar et al. analyzed the contact angle of a hydrogel composed of SA and carboxymethyl cellulose in a 1:1 ratio, reporting a contact angle value of $48.32 \pm 1.5^\circ$. Contact angle measurements were recorded using distilled water, and 3 measurements were taken to obtain the mean value.⁶⁰ Mujawar et al. conducted contact angle measurements on 3D-printed samples, including those made from SA–gelatin and SA–gelatin–*Aloe barbadensis* extract. Six contact angle measurements were taken using water as the probe liquid. The recorded water contact angle for 3D-printed SA–gelatin was $42.31 \pm 3.23^\circ$. The water contact angle reported for SA–gelatin–*Aloe barbadensis* extract was about $53.2 \pm 4.7^\circ$.⁶¹ Sisakht et al. prepared a hydrogel with a 50:50 weight ratio of SA and poly(acrylic acid). They utilized both SA and poly(acrylic acid) at a concentration of 2% wt/vol. They also analyzed the contact angle in combination with fibroblast growth factor (FGF1). Deionized water was used as the probe liquid for the contact angle analysis. Three measurements were recorded for the samples. The contact

angle reported for SA was 57.87°. In the case of the SA–poly(acrylic acid) hydrogel, the reported contact angle was 18.52°. The SA–poly(acrylic acid) hydrogel in combination with FGF1 exhibited a water contact angle of approx. 62.56°.⁶² Hosseini et al. prepared a hydrogel composed of carboxymethyl cellulose and SA. Subsequently, they combined it with simvastatin, resulting in a final form known as the carboxymethyl cellulose/SA–simvastatin hydrogel. The recorded water contact angles for carboxymethyl cellulose–SA and carboxymethyl cellulose/SA–simvastatin were $43.3 \pm 2^\circ$ and $59.1 \pm 4^\circ$, respectively. They utilized water as the probe liquid, and 3 measurements were recorded for each sample.⁶³ Table 1 presents a summary of the contact angle measurements for SA-based materials. Isa Rahim et al. specified that contact angles lower than 90° were indicative of hydrophilicity, and contact angles between 90° and 150° were indicative of hydrophobicity.⁶⁴ Chen et al. specified that a surface with a contact angle higher than 150° can be reported as superhydrophobic.⁶⁵ Choi et al. reported the use of SA for wound dressings, and the reason given in their study was its hydrophilicity.⁶⁶ Jin et al. examined the impact of hydrophilic polymers on swelling, and their study concluded that SA presented the maximum swelling among the analyzed hydrophilic polymers.⁶⁷ Feng and Wang highlighted the use of hydrogels with maximum swelling capacity for wound healing due to their ability to absorb wound exudates.⁶⁸ Singh and Pramanik reported better cell adhesion for polymeric films composed of SA and chitosan than for alginate itself, and it was also mentioned in their study that the contact angle was lower than 90°.⁶⁹

Sodium alginate wound healing

Zahid et al. indicated that the use of SA for wound healing applications is promising due to its liquid absorption.⁷⁰ The stages of wound healing have been demonstrated

in various studies and include hemostasis, inflammation, proliferation, and remodeling phases.^{71–73} Hemostasis is the first stage of wound healing, and in this phase, blood clot formation takes place.⁷⁴ Periyah et al. explained the blood coagulation mechanism in significant detail. Their study indicated the key role of platelets in thrombus formation.⁷⁵ Zhang et al. demonstrated that platelets play a vital role in vascular repair.⁷⁶ Huang et al. reported that SA can enhance platelet concentration.⁷⁷ Zhang et al. recommended SA for blood coagulation applications.⁷⁸ Chen et al. reported in their study the preparation of a hemostatic powder, which involves the presence of SA, CaCl₂, graphene oxide, and cerium nitrate. The concluding remarks in their study indicated that the hemostatic powder presented the ability to absorb water, resulting in a shorter hemostatic time. The hemostatic powder effectively decreased blood loss and enhanced the wound healing mechanism due to the presence of Ce³⁺ (which acts as a free radical scavenger).⁷⁹ Zhou et al. analyzed the blood coagulation of a gelatin sponge individually and in combination with SA, and their study summarized that blood coagulation was enhanced due to the presence of SA.⁸⁰ Wang et al. prepared a powdered composite that included SA and poly(γ -glutamic acid) (PGA). The prepared powdered composite was found to arrest or block bleeding.⁸¹ Li et al. presented a material composed of SA, silk fibroin, and thrombin. It was mentioned that the prepared material could promote hemostasis.⁸² Xie et al. prepared a hydrogel containing SA, chitosan, and oxidized dextran. Their study results showed that the prepared hydrogel reduced blood loss, which significantly supports the basis for improved hemostasis.⁸³ Khatkhat et al. mentioned that persistent inflammation was a significant reason for the slowing down or delay of the wound healing mechanism.⁸⁴ Zhang et al. reported that inflammation was a notable challenge in wound healing.⁸⁵ Liu et al. highlighted that the anti-inflammatory effect was essential for wound healing.⁸⁶ Summa et al. prepared a wound dressing material composed of SA and povidone–iodine.

Table 1. Contact angle measurements for sodium alginate (SA)-based materials

SA 200	3	Milli-Q water	$68.5 \pm 2.4^\circ$	59
SA–carboxymethyl cellulose (1:1 ratio)	3	distilled water	$48.32 \pm 1.5^\circ$	60
SA–gelatin– <i>Aloe barbadensis</i> extract	6	water	$53.2 \pm 4.7^\circ$	61
SA–poly(acrylic acid) (50:50)	3	deionized water	18.52°	62
Carboxymethyl cellulose–SA	3	water	$43.3 \pm 2^\circ$	63

The results of their study indicated that the prepared material exhibited an anti-inflammatory effect.⁸⁷ Zhao et al. formulated a composite that included low-viscosity SA combined with titanium dioxide and temozolomide nanoparticles. Their study reported the anti-inflammatory effect of the designed composite. Their research summarized that the anti-inflammatory effect was enhanced due to the prominent presence of SA.⁸⁸ Karmakar et al. initially prepared a SA and carboxymethyl cellulose composite, which was then crosslinked to a decellularized extracellular matrix. The crosslinker used in their study was calcium chloride, resulting in a final hydrogel composed of SA, carboxymethyl cellulose, and decellularized extracellular matrix. According to their research, the prepared hydrogel exhibited anti-inflammatory properties.⁶⁰ Zhang et al. prepared a blend of SA and carboxymethyl chitosan, which was then loaded with curcumin. Sr^{2+} was used for the crosslinking of the polymers. The prepared hydrogel exhibited anti-inflammatory activity, leading to an enhanced wound healing mechanism.⁸⁹ Raguvaran et al. prepared hydrogels composed of SA and gum acacia, which were incorporated with zinc oxide nanoparticles. Their research concluded that polymers in combination with zinc oxide nanoparticles tend to boost the proliferation of fibroblasts.⁹⁰ Song et al. demonstrated that fibroblast proliferation is essential to promote the wound healing mechanism.⁹¹ Harper et al. presented fibroblast migration as a component of the proliferative phase of wound healing.⁹² Kibungu et al. reported that fibroblast cells assist in the production of collagen.⁹³ AlShaali et al. also described that granulation tissue starts to form during the proliferation stage.⁹⁴ Zhu et al. utilized a hydrogel based on SA, which was incorporated with *Capparis spinosa* L., and the incorporated hydrogel exhibited biocompatibility. It also boosted cell proliferation.⁹⁵ Zhou et al. reported that enhanced cell proliferation encouraged wound healing.⁹⁶ Ma et al. synthesized a nanocomposite based on SA, polyvinyl alcohol, and graphene oxide. Their research summarized that nanocomposites may result in enhanced cell proliferation.⁹⁷ Doderio et al. prepared electrospun membranes of SA, which were crosslinked with divalent cations such as Ca^{2+} , Sr^{2+} , and Ba^{2+} . In the final step, they prepared electrospun mats of SA loaded with zinc oxide nanoparticles, which were crosslinked with Sr^{2+} ions due to excellent cell adhesion. The electrospun mat prepared in their research was recommended for wound healing applications.⁹⁸ Ding et al. utilized electrostatic spinning to synthesize a membrane based on SA and polyvinyl alcohol. Shikonin was incorporated into the electrospun membrane. It was found that the shikonin-loaded membrane boosted vascular endothelial growth factor A (VEGF-A) in the proliferative stage of wound healing.⁹⁹ Liu et al. reported shikonin as a naphthoquinone pigment useful in the preparation of shikonin-based nanomedicine.¹⁰⁰ Guo et al. highlighted *Lithospermum erythrorhizon* as the source of extraction for shikonin.¹⁰¹ Lin et al. referred

to *Lithospermum erythrorhizon* as a Chinese medicinal herb.¹⁰² Tian et al. reported the antimicrobial and anti-inflammatory properties of shikonin in their research.¹⁰³ Guo et al. mentioned the antioxidant and antithrombotic properties of shikonin.¹⁰¹ Ye et al. reported the anti-tumor effect of shikonin.¹⁰⁴ Several studies reported shikonin as an ideal option for the treatment of diabetic wounds.^{105–107} Eming et al. described VEGF-A in relation to angiogenesis in wound healing.¹⁰⁸ DiPietro presented that angiogenesis results from hypoxia.¹⁰⁹ In the case of injury, hypoxia is primarily responsible for the recruitment of hypoxia-inducible factors such as HIF-1. It supports the mechanism of angiogenesis during wound healing.¹¹⁰ Dawood et al. highlighted angiogenesis as a key feature linked to the proliferative phase of wound healing.¹¹¹ Bahadoran et al. referred to the proliferative phase by another name, the growth phase of wound healing.¹¹² Çerçi et al. utilized amoxicillin in combination with SA and polyvinyl alcohol. Polyvinyl alcohol was employed at a concentration of 12% wt/vol. Sodium alginate was utilized at a concentration of 1% wt/vol. The ratio selected for polyvinyl alcohol and SA was 2:1 (vol/vol) for the electrospinning solution, and 6.4 mg of amoxicillin trihydrate was added to the electrospinning solution. The electrospun nanofibrous mats were prepared by electrospinning. According to their research conclusions, the amoxicillin-loaded electrospun nanofibrous mat exhibited antibacterial properties. Based on their in vitro studies, the prepared nanofibrous mat loaded with amoxicillin was recommended for wound dressing applications.¹¹³ Kaur et al. reported amoxicillin as an antibiotic belonging to the penicillin class.¹¹⁴ Ayavoo et al. reported that the remodeling phase of wound healing was responsible for the conversion or transformation of fibroblasts into myofibroblasts.¹¹⁵ The remodeling phase of wound healing tends to remodel collagen III to collagen I.¹¹⁶ Yang et al. recommended a scaffold composed of SA, silk fibroin, and hyaluronic acid for extracellular matrix remodeling.¹¹⁷

Bibliometric analysis

The bibliometric analysis was conducted using the R software v. 4.5.1 (R Foundation for Statistical Computing, Vienna, Austria) and VOSviewer (<https://www.vosviewer.com/>). Initially, the “bibliometrix” package was installed, followed by loading the library (bibliometrix). The biblioshiny() function was used to open the web-based interface for bibliometrix. Bibliometrix and biblioshiny were developed by Aria and Cuccurullo.¹¹⁸ The data were obtained from Scopus, and the document search was conducted using the terms “Sodium Alginate” AND “Wound Healing”, with the search restricted to “Article title, Abstract, Keywords” from 2010 to 2025 (access date: 27 June 2025). A total of 944 documents were generated from the search

for “Sodium Alginate” AND “Wound Healing”. The annual scientific production data from Scopus for the specified search are presented in Fig. 2. The analyzed data show that the number of articles from 2018 to 2024 increased continuously, indicating the growing prominence of the presented research in recent years. The most relevant sources are depicted in Fig. 3. It represents the *International Journal of Biological Macromolecules*, the leading journal presenting pertinent articles, with 159 publications. The 2nd most relevant source was *Carbohydrate Polymers*, with 37 articles. The 3rd most relevant source was *ACS Applied Materials*

and Interfaces, with 24 articles reported. The most globally cited papers were analyzed using Scopus data, and the results are presented in Fig. 4, which shows the papers with the most significant global citation impact. The impact of local citations was also investigated, and the highest number of locally cited documents was 32, as depicted in Fig. 5. Van Eck and Waltman developed VOSviewer for bibliometric mapping.¹¹⁹ The Scopus data file was imported into VOSviewer, which helped identify relevant keywords through co-occurrence analysis. The co-occurrence analysis is presented in Fig. 6.

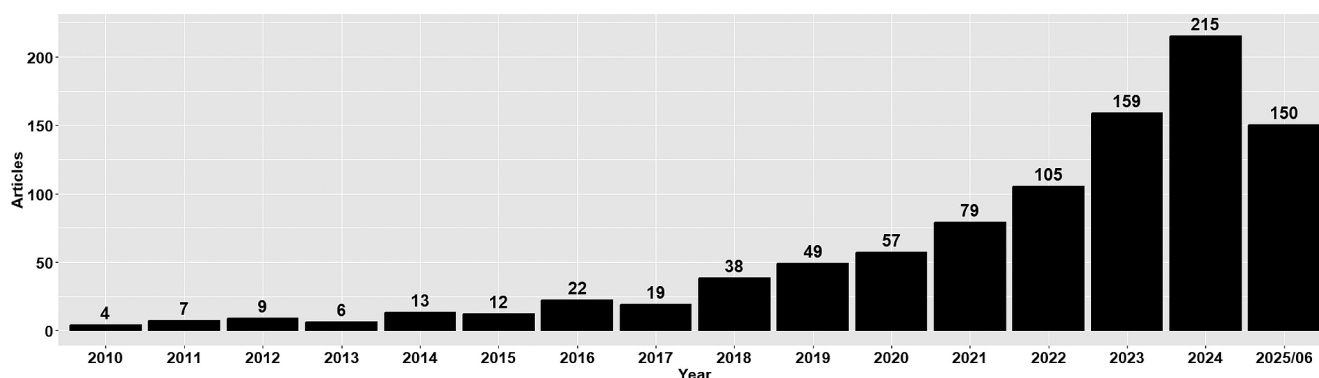


Fig. 2. Annual scientific production (“Sodium Alginate” AND “Wound Healing” search within Scopus; access date: June 27, 2025)

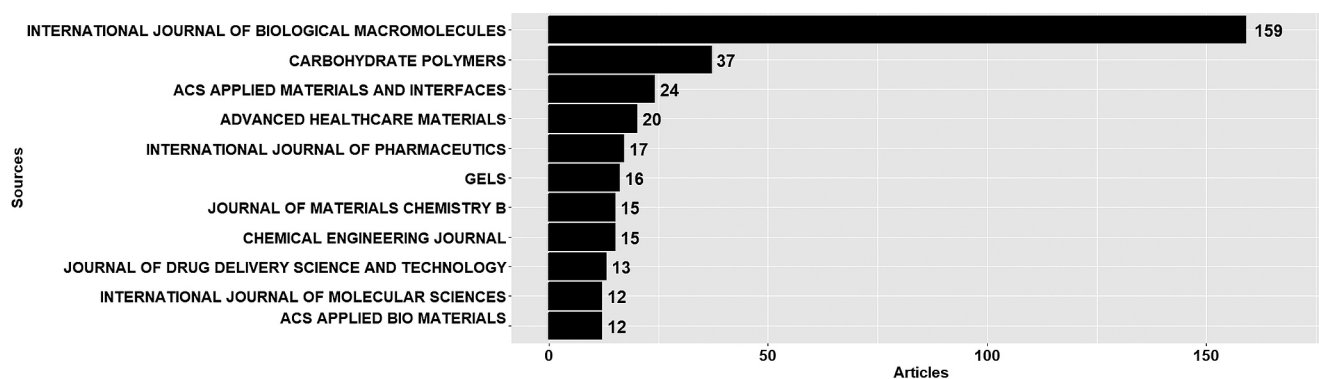


Fig. 3. Most relevant sources with the number of articles (“Sodium Alginate” AND “Wound Healing” search within Scopus; access date: June 27, 2025)

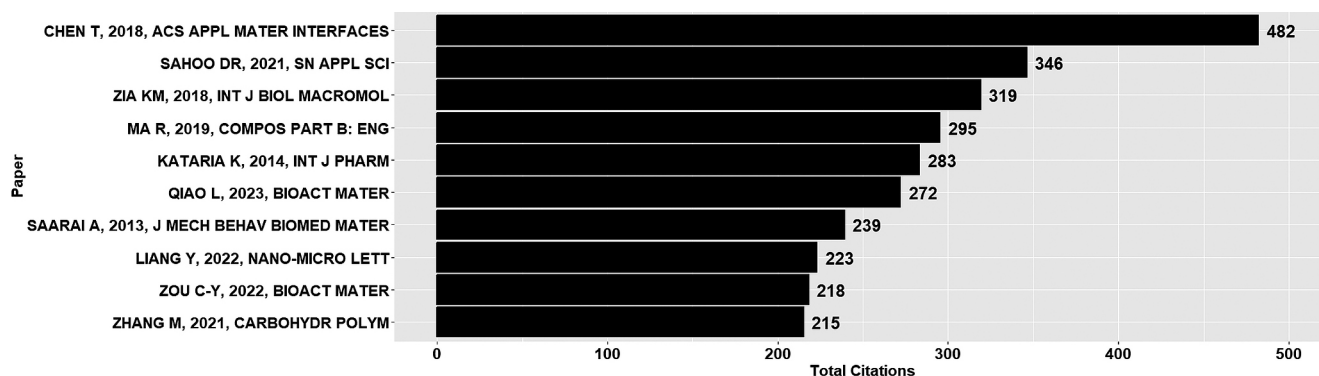


Fig. 4. Most globally cited papers (“Sodium Alginate” AND “Wound Healing” search within Scopus; access date: June 27, 2025)

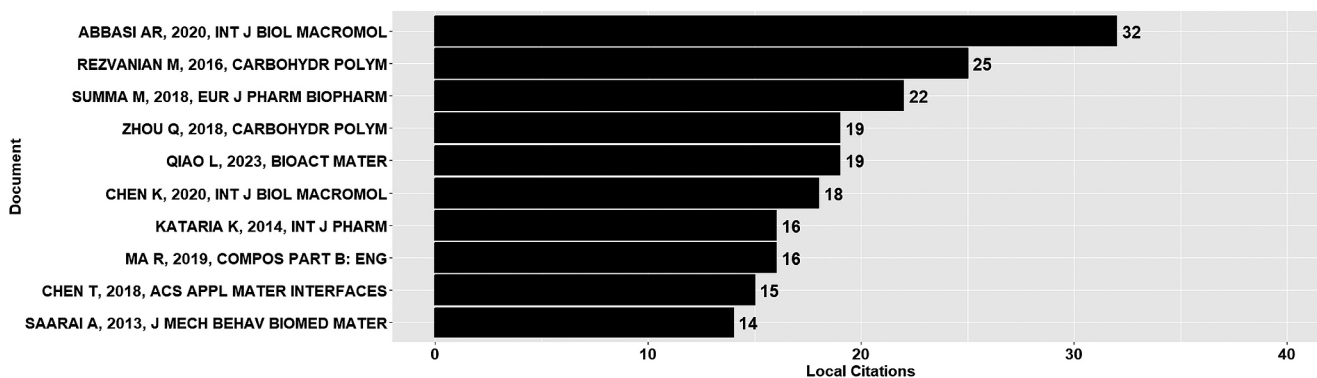


Fig. 5. Most locally cited documents (“Sodium Alginate” AND “Wound Healing” search within Scopus; access date: June 27, 2025)

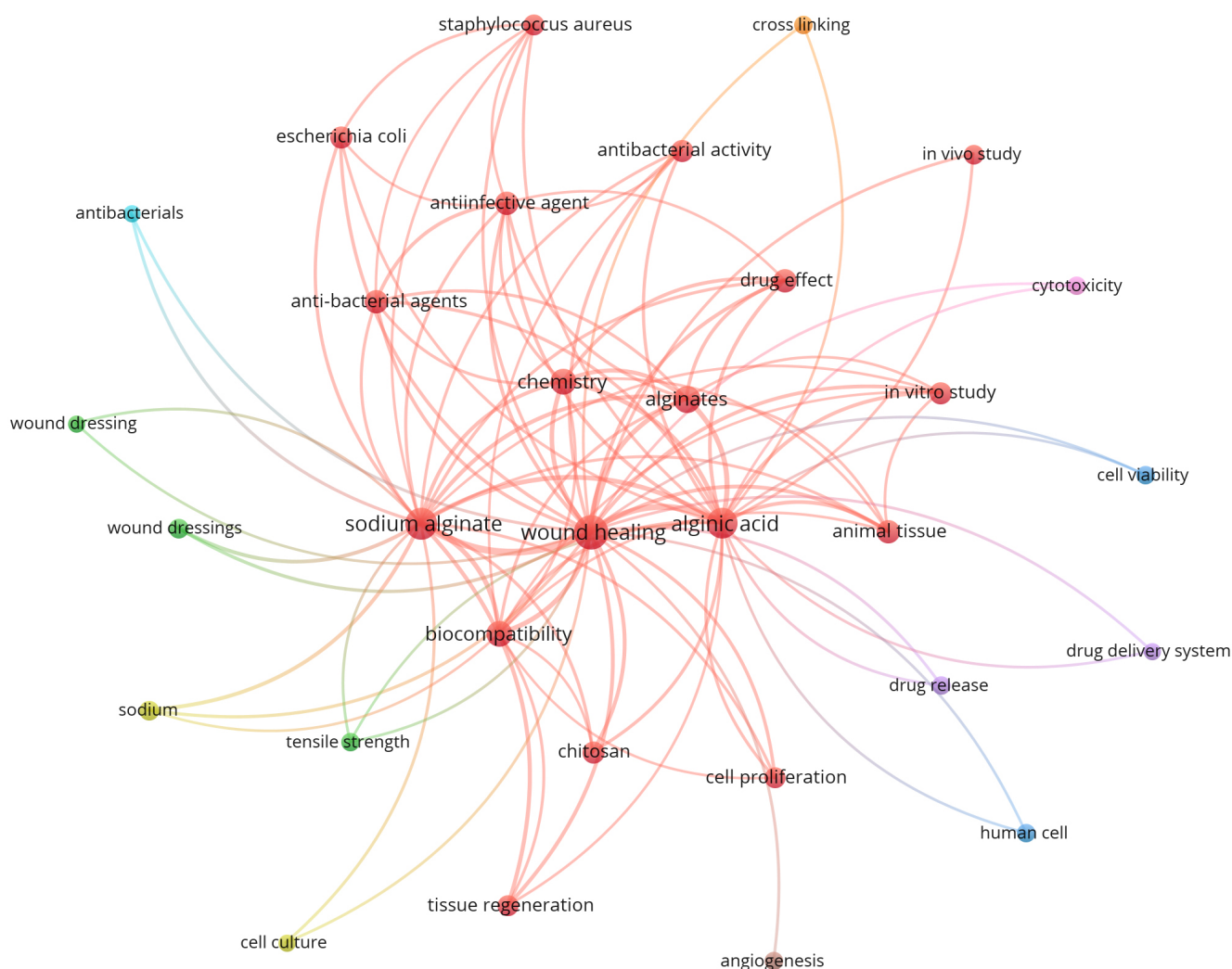


Fig. 6. Co-occurrence of keywords (“Sodium Alginate” AND “Wound Healing” search within Scopus; access date: June 27, 2025)

Conclusions and future outlook

Several factors must be considered in the wound healing mechanism, including the hydrophilicity, porosity, and biocompatibility of wound dressing materials. Sodium alginate is primarily used in hemostatic powders

for blood coagulation, as blood coagulation is the initial phase of wound healing. Wound healing is often delayed due to prolonged inflammation. Sodium alginate exhibits its significant anti-inflammatory properties, facilitating early wound healing. The proliferation phase of wound healing is the combined result of angiogenesis, fibroblast

migration, and collagen deposition. Sodium alginate can effectively promote the proliferative phase of wound healing by contributing to tissue granulation. Sodium alginate scaffolds also promote extracellular matrix remodeling. Machine learning is widely used to explore porous material structures. In the future, there is a need to develop an application programming interface (API) to facilitate data communication, particularly an API for the porosity of SA-based materials. Bibliometric analysis also revealed that open-access journals are focusing on promoting research related to SA wound healing applications. Although many papers regarding the application of SA have been published so far, it can be expected that more research articles on this material will be published in the future, as many research groups worldwide are working on alginate and its combinations with other macromolecules.

ORCID iDs

Muhammad Tahir  0000-0001-8436-1126
 Simone Pettineo  0000-0002-5812-5565
 Silvia Vicini  0000-0001-9369-1522
 Maila Castellano  0000-0002-0930-7522
 Marina Alloisio  0000-0002-6155-2912
 Ketul Popat  0000-0002-2417-7789
 Harishkumar Madhyastha  0000-0002-7143-4821
 Kaoru Ohe  0009-0005-6343-568X
 Kentaro Sakai  0000-0002-5811-8795
 Alina Sionkowska  0000-0002-1551-2725

References

- Raina N, Rani R, Pahwa R, Gupta M. Biopolymers and treatment strategies for wound healing: An insight view. *Int J Polym Mater Polym Biomater*. 2022;71(5):359–375. doi:10.1080/00914037.2020.1838518
- Jabeen N, Atif M. Polysaccharides based biopolymers for biomedical applications: A review. *Polym Adv Techs*. 2024;35(1):e6203. doi:10.1002/pat.6203
- Thakur VK, Thakur MK, eds. *Functional Biopolymers*. Cham, Switzerland: Springer International Publishing; 2018. doi:10.1007/978-3-319-66417-0
- Nitta S, Numata K. Biopolymer-based nanoparticles for drug/gene delivery and tissue engineering. *Int J Mol Sci*. 2013;14(1):1629–1654. doi:10.3390/ijms14011629
- Ahmed HB, Mikhail MM, El-Shahat M, Emam HE. Clustering of carbon quantum dots from polysaccharides (cellulose, alginate, chitosan) versus heterocyclic compounds: Synthesis, characterization and medical applications. *Carbohydr Polym Technol Appl*. 2025;9:100738. doi:10.1016/j.carpta.2025.100738
- Shi C, Xu R, Hu W, et al. Fabrication of alginate dialdehyde-gelatin crosslinked hydrogels incorporated with cinnamaldehyde nanoparticles for meat preservation. *Int J Biol Macromol*. 2025;298:140063. doi:10.1016/j.ijbiomac.2025.140063
- Kulkarni VS, Butte KD, Rathod SS. Natural polymers—a comprehensive review. *Int J Res Pharm Biomed Sci*. 2012;3(4):1597–1613. Available from: https://www.researchgate.net/profile/Vishakha-gajrekulkarni/publication/236217541_Natural_Polymers-_A_comprehensive_Review/links/595b4913a6fdcc36b4daa3f7/Natural-Polymers-A-comprehensive-Review.pdf
- Al-Roujayee AS, Hilaj E, Deepak A, et al. Alginate-based systems: Advancements in drug delivery and wound healing. *Int J Polym Mater Polym Biomater*. 2025;74(9):846–874. doi:10.1080/00914037.2024.2375343
- Lee KY, Mooney DJ. Alginate: Properties and biomedical applications. *Prog Polym Sci*. 2012;37(1):106–126. doi:10.1016/j.progpolymsci.2011.06.003
- Abourehab MAS, Rajendran RR, Singh A, et al. Alginate as a promising biopolymer in drug delivery and wound healing: A review of the state-of-the-art. *Int J Mol Sci*. 2022;23(16):9035. doi:10.3390/ijms23169035
- Saberian M, Safari Roudsari R, Haghshenas N, Roustaa A, Alizadeh S. How the combination of alginate and chitosan can fabricate a hydrogel with favorable properties for wound healing. *Heliyon*. 2024;10(11):e32040. doi:10.1016/j.heliyon.2024.e32040
- Liang X, Huang C, Liu H, et al. Natural hydrogel dressings in wound care: Design, advances, and perspectives. *Chin Chem Lett*. 2024;35(10):109442. doi:10.1016/j.ccllet.2023.109442
- Salarvand S, Jalali SAH, Mahboobi Soofiani N, Allafchian A. Amine modified sodium alginate: Synthesis, characterization and in vivo evaluation in rainbow trout (*Oncorhynchus mykiss*). *Carbohydr Polym Technol Appl*. 2025;9:100699. doi:10.1016/j.carpta.2025.100699
- Yu P, Zhong W. Hemostatic materials in wound care. *Burns Trauma*. 2021;9:tkab019. doi:10.1093/burnst/tkab019
- Wang W, Liu M, Shafiq M, et al. Synthesis of oxidized sodium alginate and its electrospun bio-hybrids with zinc oxide nanoparticles to promote wound healing. *Int J Biol Macromol*. 2023;232:123480. doi:10.1016/j.ijbiomac.2023.123480
- Dodero A, Alberti S, Gaggero G, et al. An up-to-date review on alginate nanoparticles and nanofibers for biomedical and pharmaceutical applications. *Adv Mater Interfaces*. 2021;8(22):2100809. doi:10.1002/admi.202100809
- Dodero A, Alloisio M, Castellano M, Vicini S. Multilayer alginate-poly-caprolactone electrospun membranes as skin wound patches with drug delivery abilities. *ACS Appl Mater Interfaces*. 2020;12(28):31162–31171. doi:10.1021/acsami.0c07352
- Phùng TTT, Đinh HN, Ureña M, et al. Sodium alginate as a promising encapsulating material for extremely-oxygen sensitive probiotics. *Food Hydrocoll*. 2025;160:110857. doi:10.1016/j.foodhyd.2024.110857
- Yan P, Lan W, Xie J. Modification on sodium alginate for food preservation: A review. *Trends Food Sci Technol*. 2024;143:104217. doi:10.1016/j.tifs.2023.104217
- Astaneh ME, Hashemzadeh A, Fereydouni N. Recent advances in sodium alginate-based dressings for targeted drug delivery in the context of diabetic wound healing. *J Mater Chem B*. 2024;12(40):10163–10197. doi:10.1039/D4TB01049C
- Luan Q, Wang Y, Chen Y, Chen H. Review on improvement of physicochemical properties of sodium alginate-based edible films. *J Food Sci*. 2025;90(2):e70016. doi:10.1111/1750-3841.70016
- Yang JS, Xie YJ, He W. Research progress on chemical modification of alginate: A review. *Carbohydr Polym*. 2011;84(1):33–39. doi:10.1016/j.carbpol.2010.11.048
- Belalia F, Djelali NE. Rheological properties of sodium alginate solutions. *Rev Roum Chim*. 2014;59(2):135–145. Accessed July 3, 2026. <https://revroum.lew.ro/wp-content/uploads/2014/2/Art%2008.pdf>
- Karvinen J, Kellomäki M. Design aspects and characterization of hydrogel-based bioinks for extrusion-based bioprinting. *Bioprinting*. 2023;32:e00274. doi:10.1016/j.bprint.2023.e00274
- Badawi NM, Batoo KM, Subramaniam R, et al. Highly conductive and reusable cellulose hydrogels for supercapacitor applications. *Micro-machines*. 2023;14(7):1461. doi:10.3390/mi14071461
- Pongjanyakul T. Alginate–magnesium aluminum silicate films: Importance of alginate block structures. *Int J Pharm*. 2009;365(1–2):100–108. doi:10.1016/j.ijpharm.2008.08.025
- Pathak TS, Kim JS, Lee SJ, Baek DJ, Paeng KJ. Preparation of alginate acid and metal alginate from algae and their comparative study. *J Polym Environ*. 2008;16(3):198–204. doi:10.1007/s10924-008-0097-4
- Bojorges H, López-Rubio A, Martínez-Abad A, Fabra MJ. Overview of alginate extraction processes: Impact on alginate molecular structure and techno-functional properties. *Trends Food Sci Technol*. 2023;140:104142. doi:10.1016/j.tifs.2023.104142
- Saji S, Hebden A, Goswami P, Du C. A brief review on the development of alginate extraction process and its sustainability. *Sustainability*. 2022;14(9):5181. doi:10.3390/su14095181
- Pawar SN, Edgar KJ. Alginate derivatization: A review of chemistry, properties and applications. *Biomaterials*. 2012;33(11):3279–3305. doi:10.1016/j.biomaterials.2012.01.007
- Abka-khajouei R, Tounsi L, Shahabi N, Patel AK, Abdelkafi S, Michaud P. Structures, properties and applications of alginates. *Mar Drugs*. 2022;20(6):364. doi:10.3390/md20060364
- Da Silva CM, Reis RL, Correlo VM, Jahno VD. The efficient role of sodium alginate-based biodegradable dressings for skin wound healing application: A systematic review. *J Biomater Sci Polym Ed*. 2024;35(3):397–414. doi:10.1080/09205063.2023.2289247
- Sinha A, Georgoulas A, Crua C, et al. Exploring exudate absorp-

- tion via sessile droplet dynamics in porous wound dressings. *Exp Therm Fluid Sci.* 2025;163:111408. doi:10.1016/j.expthermflusc.2025.111408
34. Liu S, Li H, Tang B, Bi S, Li L. Scaling law and microstructure of alginate hydrogel. *Carbohydr Polym.* 2016;135:101–109. doi:10.1016/j.carbpol.2015.08.086
 35. Androniceanu M. Efficiency and prediction in human resource management using Python modules. *Theor Empir Res Urban Manag.* 2025;20(1):88–103. Available from: <https://um.ase.ro/v20i1/5.pdf>
 36. Reis E, Gralha C, Monteiro MP. Surveying communities of users of MATLAB and clone languages. *J Comp Lang.* 2022;73:101170. doi:10.1016/j.cola.2022.101170
 37. Jenkins D, Salhadar K, Ashby G, et al. PoreScript: Semi-automated pore size algorithm for scaffold characterization. *Bioact Mater.* 2022;13:1–8. doi:10.1016/j.bioactmat.2021.11.006
 38. Polez RT, Ajiboye MA, Österberg M, Horn MM. Chitosan hydrogels enriched with bioactive phloroglucinol for controlled drug diffusion and potential wound healing. *Int J Biol Macromol.* 2024;265:130808. doi:10.1016/j.ijbiomac.2024.130808
 39. Gostick J, Khan Z, Tranter T, et al. PoreSpy: A Python toolkit for quantitative analysis of porous media images. *J Open Source Soft.* 2019;4(37):1296. doi:10.21105/joss.01296
 40. Negut I, Bitu B. Exploring the potential of artificial intelligence for hydrogel development: A short review. *Gels.* 2023;9(11):845. doi:10.3390/gels9110845
 41. Zhang Q, Yang LT, Chen Z, Li P. A survey on deep learning for big data. *Information Fusion.* 2018;42:146–157. doi:10.1016/j.inffus.2017.10.006
 42. El Naqa I, Murphy MJ. What are machine and deep learning? In: El Naqa I, Murphy MJ, eds. *Machine and Deep Learning in Oncology, Medical Physics and Radiology.* Cham, Switzerland: Springer International Publishing; 2022:3–15. doi:10.1007/978-3-030-83047-2_1
 43. Zhang B, Shi H, Wang H. Machine learning and AI in cancer prognosis, prediction, and treatment selection: A critical approach. *J Multi-discip Healthc.* 2023;16:1779–1791. doi:10.2147/JMDH.S410301
 44. Chen Y, You W, Ou L, Tang H. A review of machine learning techniques for urban resilience research: The application and progress of different machine learning techniques in assessing and enhancing urban resilience. *Syst Soft Comput.* 2025;7:200269. doi:10.1016/j.sasc.2025.200269
 45. Martínez Torres J, Iglesias Comesaña C, García-Nieto PJ. Review: Machine learning techniques applied to cybersecurity. *Int J Mach Learn Cyber.* 2019;10(10):2823–2836. doi:10.1007/s13042-018-00906-1
 46. Rashidi HH, Tran NK, Betts EV, Howell LP, Green R. Artificial intelligence and machine learning in pathology: The present landscape of supervised methods. *Acad Pathol.* 2019;6:2374289519873088. doi:10.1177/2374289519873088
 47. Dike HU, Zhou Y, Deveerasetty KK, Wu Q. Unsupervised learning based on artificial neural network: A review. In: *2018 IEEE International Conference on Cyborg and Bionic Systems (CBS).* Shenzhen, China: IEEE; 2018:322–327. doi:10.1109/CBS.2018.8612259
 48. Nayeri ZM, Ghafarian T, Javadi B. Application placement in Fog computing with AI approach: Taxonomy and a state of the art survey. *J Netw Comp Appl.* 2021;185:103078. doi:10.1016/j.jnca.2021.103078
 49. Vaish R, Dwivedi UD, Tewari S, Tripathi SM. Machine learning applications in power system fault diagnosis: Research advancements and perspectives. *Eng Appl Artif Intelligence.* 2021;106:104504. doi:10.1016/j.engappai.2021.104504
 50. Saberian M, Seyedjafari E, Zargar SJ, Mahdavi FS, Sanaei-rad P. Fabrication and characterization of honey and aloe vera for wound dressing applications. *J Appl Polym Sci.* 2021;138(47):51398. doi:10.1002/app.51398
 51. Olevsky LM, Jacques MG, Hixon KR. PoreVision: A program for enhancing efficiency and accuracy in SEM pore analyses of gels and other porous materials. *Gels.* 2025;11(2):132. doi:10.3390/gels11020132
 52. Shkarin R, Shkarin A, Shkarina S, et al. Quanfima: An open source Python package for automated fiber analysis of biomaterials. *PLoS One.* 2019;14(4):e0215137. doi:10.1371/journal.pone.0215137
 53. Rouhollahi A, Ilegbusi O, Foroosh H. Segmentation and pore structure estimation in SEM images of tissue engineering scaffolds using genetic algorithm. *Ann Biomed Eng.* 2021;49(3):1033–1045. doi:10.1007/s10439-020-02638-2
 54. Rouhollahi A. Integration of computational fluid dynamics and machine learning for modeling scaffold pore structure for tissue engineering [dissertation]. Orlando, FL: University of Central Florida; 2019. Accessed July 3, 2026. <https://stars.library.ucf.edu/etd/6880>
 55. Nair M, Shepherd JH, Best SM, Cameron RE. MicroCT analysis of connectivity in porous structures: Optimizing data acquisition and analytical methods in the context of tissue engineering. *J R Soc Interface.* 2020;17(165):20190833. doi:10.1098/rsif.2019.0833
 56. Karaca I, Aldemir Dikici B. Quantitative evaluation of the pore and window sizes of tissue engineering scaffolds on scanning electron microscope images using deep learning. *ACS Omega.* 2024;9(23):24695–24706. doi:10.1021/acsomega.4c01234
 57. Yuan Y, Lee TR. Contact angle and wetting properties. In: Bracco G, Holst B, eds. *Surface Science Techniques.* Vol. 51. Springer Series in Surface Sciences. Berlin–Heidelberg, Germany: Springer Berlin Heidelberg; 2013:3–34. doi:10.1007/978-3-642-34243-1_1
 58. Kumar G, Prabhu KN. Review of non-reactive and reactive wetting of liquids on surfaces. *Adv Colloid Interface Sci.* 2007;133(2):61–89. doi:10.1016/j.cis.2007.04.009
 59. Xie H, Li X, Lv G, et al. Effect of surface wettability and charge on protein adsorption onto implantable alginate–chitosan–alginate microcapsule surfaces. *J Biomed Mater Res.* 2010;92A(4):1357–1365. doi:10.1002/jbm.a.32437
 60. Karmakar R, Dixit M, Eswar K, et al. Enhanced wound healing properties by sodium alginate–carboxymethyl cellulose hydrogel enriched with decellularized amniotic membrane. *Eur J Pharm Biopharm.* 2025;207:114621. doi:10.1016/j.ejpb.2024.114621
 61. Mujawar SS, Arbade GK, Bisht N, et al. 3D printed *Aloe barbadensis* loaded alginate–gelatin hydrogel for wound healing and scar reduction: In vitro and in vivo study. *Int J Biol Macromol.* 2025;296:139745. doi:10.1016/j.ijbiomac.2025.139745
 62. Sisakht MM, Gholizadeh F, Shahravi Z, Doust-Vaghe YK, Nilfroushzadeh MA, Amirkhani MA. Sodium alginate/poly(acrylic acid) hydrogel composite, potential carrier for fibroblast growth factor 1 (FGF1) delivery. *Chem Biodivers.* 2025;22(2):e202401738. doi:10.1002/cbdv.202401738
 63. Hosseini SMR, Heydari P, Namnabat M, et al. Carboxymethyl cellulose/sodium alginate hydrogel with anti-inflammatory capabilities for accelerated wound healing: In vitro and in vivo study. *Eur J Pharmacol.* 2024;976:176671. doi:10.1016/j.ejphar.2024.176671
 64. Isa Rahim M, Aqida SN, Salwani MS, Ahmad Syarizan S. Enhancing surface hydrophobicity of AISI 304 stainless steel via laser texturing. *J Phys Conf Ser.* 2025;2933(1):012006. doi:10.1088/1742-6596/2933/1/012006
 65. Chen S, Li S, Ye Z, et al. Superhydrophobic and superhydrophilic polyurethane sponge for wound healing. *Chem Eng J.* 2022;446:136985. doi:10.1016/j.cej.2022.136985
 66. Choi YK, Din FU, Kim DW, et al. Amniotic membrane extract-loaded double-layered wound dressing: Evaluation of gel properties and wound healing. *Drug Dev Ind Pharm.* 2014;40(7):852–859. doi:10.3109/03639045.2013.788015
 67. Jin SG, Yousaf AM, Kim KS, et al. Influence of hydrophilic polymers on functional properties and wound healing efficacy of hydrocolloid based wound dressings. *Int J Pharm.* 2016;501(1–2):160–166. doi:10.1016/j.ijpharm.2016.01.044
 68. Feng W, Wang Z. Tailoring the swelling–shrinkable behavior of hydrogels for biomedical applications. *Adv Sci.* 2023;10(28):2303326. doi:10.1002/adv.202303326
 69. Singh AK, Pramanik K. Fabrication and investigation of physicochemical and biological properties of 3D printed sodium alginate–chitosan blend polyelectrolyte complex scaffold for bone tissue engineering application. *J Appl Polym Sci.* 2023;140(12):e53642. doi:10.1002/app.53642
 70. Zahid M, Lodhi M, Afzal A, et al. Development of hydrogels with the incorporation of *Raphanus sativus* L. seed extract in sodium alginate for wound-healing application. *Gels.* 2021;7(3):107. doi:10.3390/gels7030107
 71. Almadani YH, Vorstenbosch J, Davison PG, Murphy AM. Wound healing: A comprehensive review. *Semin Plast Surg.* 2021;35(3):141–144. doi:10.1055/s-0041-1731791
 72. Fahimirad S, Fattahi F, Hatami M, Shabani S, Ghorbanpour M. Nanotechnology-based biotherapeutics for physiological wound healing phases. *Ind Crops Products.* 2025;226:120608. doi:10.1016/j.indcrop.2025.120608
 73. Sadeghi M, Moghaddam A, Amiri AM, et al. Improving the wound healing process: Pivotal role of mesenchymal stromal/stem cells and immune cells. *Stem Cell Rev Rep.* 2025;21(3):680–697. doi:10.1007/s12015-025-10849-0

74. Ukaegbu K, Allen E, Svoboda KKH. Reactive oxygen species and antioxidants in wound healing: Mechanisms and therapeutic potential. *Int Wound J.* 2025;22(5):e70330. doi:10.1111/iwj.70330
75. Periyah MH, Halim AS, Mat Saad AZ. Mechanism action of platelets and crucial blood coagulation pathways in hemostasis. *Int J Hematol Oncol Stem Cell Res.* 2017;11(4):319–327. PMID:29340130. PMID:PMCS767294.
76. Zhang P, Zu R, Zhang X, et al. Unveiling the “Dark Matter” of platelet involvement in tumor microenvironment. *J Pharm Anal.* 2025;15(9):101218. doi:10.1016/j.jpba.2025.101218
77. Huang H, Chen H, Wang X, et al. Degradable and bioadhesive alginate-based composites: An effective hemostatic agent. *ACS Biomater Sci Eng.* 2019;5(10):5498–5505. doi:10.1021/acsbiomaterials.9b01120
78. Zhang J, Zhang S, Liu C, et al. Photopolymerized multifunctional sodium alginate-based hydrogel for antibacterial and coagulation dressings. *Int J Biol Macromol.* 2024;260:129428. doi:10.1016/j.ijbiomac.2024.129428
79. Chen A, Wu L, Wang K, et al. Facile synthesis of rapid hemostatic powder based on sodium alginate for promoting hemostasis and wound healing. *Int J Biol Macromol.* 2025;308:142728. doi:10.1016/j.ijbiomac.2025.142728
80. Zhou J, Li M, Hui Y, et al. Hemostatic sponge based on easily prepared crosslinked gelatin and sodium alginate for wound healing. *J Mater Sci.* 2024;59(19):8408–8426. doi:10.1007/s10853-024-09539-y
81. Wang Y, Wang P, Ji H, Ji G, Wang M, Wang X. Analysis of safety and effectiveness of sodium alginate/poly(γ -glutamic acid) microspheres for rapid hemostasis. *ACS Appl Biomater.* 2021;4(8):6539–6548. doi:10.1021/acsabm.1c00671
82. Li Y, Li M, Li C, et al. A sodium alginate – silk fibroin biosponge loaded with thrombin: Effective hemostasis and wound healing. *Heliyon.* 2024;10(6):e28047. doi:10.1016/j.heliyon.2024.e28047
83. Xie M, Zeng Y, Wu H, Wang S, Zhao J. Multifunctional carboxymethyl chitosan/oxidized dextran/sodium alginate hydrogels as dressing for hemostasis and closure of infected wounds. *Int J Biol Macromol.* 2022;219:1337–1350. doi:10.1016/j.ijbiomac.2022.08.166
84. Khattak S, Ullah I, Sohail M, et al. Endogenous/exogenous stimuli-responsive smart hydrogels for diabetic wound healing. *Aggregate.* 2025;6(2):e688. doi:10.1002/agt2.688
85. Zhang X, Li R, Li S, et al. Tri-network PVA/chitosan/gelatin hydrogel modified by tannic acid with self-healing, adhesive and anti-inflammatory properties to accelerate wound healing. *Int J Biol Macromol.* 2025;308:142280. doi:10.1016/j.ijbiomac.2025.142280
86. Liu H, Qin S, Zhang H, et al. Silk sericin-based ROS-responsive oxygen generating microneedle platform promotes angiogenesis and decreases inflammation for scarless diabetic wound healing. *Adv Funct Mater.* 2025;35(7):2404461. doi:10.1002/adfm.202404461
87. Summa M, Russo D, Penna I, et al. A biocompatible sodium alginate/povidone iodine film enhances wound healing. *Eur J Pharm Biopharm.* 2018;122:17–24. doi:10.1016/j.ejpb.2017.10.004
88. Zhao J, Yao L, Nie S, Xu Y. Low-viscosity sodium alginate combined with TiO₂ nanoparticles for improving neuroblastoma treatment. *Int J Biol Macromol.* 2021;167:921–933. doi:10.1016/j.ijbiomac.2020.11.048
89. Zhang X, Liu Y, Wang Z, et al. pH-responsive and self-adaptive injectable sodium alginate/carboxymethyl chitosan hydrogel accelerates infected wound healing by bacteriostasis and immunomodulation. *Carbohydr Polym.* 2025;354:123322. doi:10.1016/j.carbpol.2025.123322
90. Raguvaran R, Manuja BK, Chopra M, et al. Sodium alginate and gum acacia hydrogels of ZnO nanoparticles show wound healing effect on fibroblast cells. *Int J Biol Macromol.* 2017;96:185–191. doi:10.1016/j.ijbiomac.2016.12.009
91. Song J, Zeng L, Ye Z, et al. The promoting effect of *Balanus albicostatus* cement protein 19k (Balcp19k) on wound healing by regulating fibroblast migration and relieving early-stage inflammation responses. *Int J Biol Macromol.* 2025;289:138781. doi:10.1016/j.ijbiomac.2024.138781
92. Harper D, Young A, McNaught CE. The physiology of wound healing. *Surgery (Oxford).* 2014;32(9):445–450. doi:10.1016/j.mpsur.2014.06.010
93. Kibungu C, Kondiah PPD, Kumar P, Choonara YE. Recent advances in chitosan and alginate-based hydrogels for wound healing application. *Front Mater.* 2021;8:681960. doi:10.3389/fmats.2021.681960
94. AlShaali S, Atieh M, Hakam A, Alsabeeha N, Shah M. Effects of hyaluronic acid gel on initial wound healing following tooth extraction and crown lengthening procedures: A retrospective analysis. *Clin Cosmet Investig Dent.* 2025;17:225–236. doi:10.2147/CCIDE.S513987
95. Zhu P, Wu J, Chang Z, et al. Sodium alginate hydrogel loaded with *Capparis spinosa* L. extract for antimicrobial and antioxidant wound dressing applications. *Int J Biol Macromol.* 2025;289:138883. doi:10.1016/j.ijbiomac.2024.138883
96. Zhou C, Jiang T, Liu S, et al. AgNPs loaded adenine-modified chitosan composite POSS-PEG hybrid hydrogel with enhanced antibacterial and cell proliferation properties for promotion of infected wound healing. *Int J Biol Macromol.* 2024;267:131575. doi:10.1016/j.ijbiomac.2024.131575
97. Ma R, Wang Y, Qi H, et al. Nanocomposite sponges of sodium alginate/graphene oxide/polyvinyl alcohol as potential wound dressing: In vitro and in vivo evaluation. *Composites Part B Engineering.* 2019;167:396–405. doi:10.1016/j.compositesb.2019.03.006
98. Dodero A, Scarfi S, Pozzolini M, Vicini S, Alloisio M, Castellano M. Alginate-based electrospun membranes containing ZnO nanoparticles as potential wound healing patches: Biological, mechanical, and physicochemical characterization. *ACS Appl Mater Interfaces.* 2020;12(3):3371–3381. doi:10.1021/acsami.9b17597
99. Ding C, Yang J, Wang N, et al. Sodium alginate/polyvinyl alcohol nanofibers loaded with shikonin for diabetic wound healing: In vivo and in vitro evaluation. *Int J Biol Macromol.* 2024;262:129937. doi:10.1016/j.ijbiomac.2024.129937
100. Liu Z, Sun R, Huang Y, et al. The preparation of shikonin-based nanomedicine with robust GSH-responsiveness. *Coll Surf A Physicochem Eng Aspects.* 2025;705:135557. doi:10.1016/j.colsurfa.2024.135557
101. Guo C, He J, Song X, et al. Pharmacological properties and derivatives of shikonin: A review in recent years. *Pharmacol Res.* 2019;149:104463. doi:10.1016/j.phrs.2019.104463
102. Lin W, Wang X, Zhuang T, et al. Lithospermum erythrorhizon polysaccharide alleviates obesity via gut microbiota-mediated reprogramming of bile acid and short-chain fatty acid metabolism. *Int J Biol Macromol.* 2025;323:147082. doi:10.1016/j.ijbiomac.2025.147082
103. Tian M, Wu J, Du Q, et al. Revealing the mechanisms of shikonin against diabetic wounds: A combined network pharmacology and in vitro investigation. *J Diabetes Res.* 2025;2025(1):4656485. doi:10.1155/jdr/4656485
104. Ye X, Wu X, Lian S, et al. Recent advances of shikonin in the molecular mechanisms of anticancer, anti-inflammation and immunoregulation. *Am J Chin Med.* 2025;53(4):1093–1118. doi:10.1142/S0192415X25500417
105. Monga A, Kaur K, Gasso S, et al. Fabrication of starch nanofibrous scaffolds co-loaded with shikonin microsponges for accelerated diabetic wound healing: Deciphering in vitro insights and in vivo realities. *BioNanoScience.* 2025;15(3):319. doi:10.1007/s12668-025-01926-2
106. Luo B, Ding X, Hu Y, et al. Shikonin hastens diabetic wound healing by inhibiting M1 macrophage polarisation through the MAPK signaling pathway. *Mol Immunol.* 2025;177:73–84. doi:10.1016/j.molimm.2024.12.002
107. Cen LS, Cao Y, Zhou YM, Guo J, Xue JW. Shikonin protects mitochondria through the NFAT5/AMPK pathway for the treatment of diabetic wounds. *World J Diabetes.* 2024;15(12):2338–2352. doi:10.4239/wjdv15.i12.2338
108. Eming S, Brachvogel B, Odorisio T, Koch M. Regulation of angiogenesis: Wound healing as a model. *Prog Histochem Cytochem.* 2007;42(3):115–170. doi:10.1016/j.proghi.2007.06.001
109. DiPietro LA. Angiogenesis and wound repair: When enough is enough. *J Leukoc Biol.* 2016;100(5):979–984. doi:10.1189/jlb.4MR0316-102R
110. Okonkwo U, DiPietro L. Diabetes and wound angiogenesis. *Int J Mol Sci.* 2017;18(7):1419. doi:10.3390/ijms18071419
111. Dawood HZ, Ara C, Asmatullah, Jabeen S, Islam A, Ghauri ZH. Chitosan/fibroin biopolymer-based hydrogels for potential angiogenesis in developing chicks and accelerated wound healing in mice. *Biopolymers.* 2025;116(1):e23633. doi:10.1002/bip.23633
112. Bahadoran Z, Mirmiran P, Hosseinpahanah F, Kashfi K, Ghasemi A. Nitric oxide-based treatments improve wound healing associated with diabetes mellitus. *Med Gas Res.* 2025;15(1):23–35. doi:10.4103/mgr.MEDGASRES-D-24-00020
113. Çerçi A, Demir ES, Karaca E, Güzel ÇB, Osman B. Preparation and characterization of amoxicillin-loaded polyvinyl alcohol/sodium alginate nanofibrous mat: Drug release properties, antibacterial activity, and cytotoxicity. *Arab J Sci Eng.* 2025;50(1):77–91. doi:10.1007/s13369-024-09075-6

114. Kaur SP, Rao R, Nanda S. Amoxicillin: a broad spectrum antibiotic. *Int J Pharm Pharm Sci.* 2011;3(3):30–37. Available from: <https://innovare-academics.in/journal/ijpps/Vol3Issue3/2249.pdf>
115. Ayavoo T, Murugesan K, Gnanasekaran A. Roles and mechanisms of stem cell in wound healing. *Stem Cell Investig.* 2021;8:4. doi:10.21037/sci-2020-027
116. Ud-Din S, Bayat A. Non-invasive objective devices for monitoring the inflammatory, proliferative and remodelling phases of cutaneous wound healing and skin scarring. *Exp Dermatol.* 2016;25(8):579–585. doi:10.1111/exd.13027
117. Yang W, Xu H, Lan Y, et al. Preparation and characterisation of a novel silk fibroin/hyaluronic acid/sodium alginate scaffold for skin repair. *Int J Biol Macromol.* 2019;130:58–67. doi:10.1016/j.ijbiomac.2019.02.120
118. Aria M, Cuccurullo C. bibliometrix: An R-tool for comprehensive science mapping analysis. *J Informetrics.* 2017;11(4):959–975. doi:10.1016/j.joi.2017.08.007
119. Van Eck NJ, Waltman L. Software survey: VOSviewer, a computer program for bibliometric mapping. *Scientometrics.* 2010;84(2):523–538. doi:10.1007/s11192-009-0146-3

